Chemical Constituents of Phyllanthus reticulatus

by Ming-Sheng Lan^a), Jian-Xiong Ma^a)^b)^c), Chang-Heng Tan^{*b}), Song Wei^c), and Da-Yuan Zhu^b)

^a) Guangxi Institute of Medicinal Plant, Nanning 530023, P. R. China
^b) Department of Natural Medicinal Chemistry, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, 555 ZuChongZhi Road, Shanghai 201203, P. R. China (phone and fax: +86-21-50806728; e-mail: chtan@mail.shcnc.ac.cn)

^c) Guangxi Traditional Chinese Medical University, Nanning 530001, P. R. China

A new purine derivative, 3-(3-methylbut-2-en-1-yl)isoguanine (1), and a new cleistanthane-type diterpenoid glucoside, 19-hydroxyspruceanol 19-O- β -D-glucopyranoside (2), together with eight known compounds were isolated from the whole plant of *Phyllanthus reticulatus*. The structures were elucidated by chemical and spectroscopic methods.

Introduction. – Phyllanthus reticulatus POIR. (Euphorbiaceae) is a folk medicine used for anti-inflammation and as analgetic, and for treatment of rheumatism in the Guangxi Zhuang national area of China [1]. It is a bushy shrub distributed widely in the tropics, from tropical Africa to India, China, and South-East Asia, and south to Queensland (northern Australia) [2]. Some reports have demonstrated that the extracts of this plant had antiplasmodial [3], antidiabetic [4], antimicrobial and cytotoxic [5], and hepatoprotective [6] bioactivities. Few studies on this plant revealed various chemical constituents such as triterpenoids, phytosterols, coumarin [5], flavonoids and phenols [7]. Our phytochemical investigation of the 75% EtOH extract of the title plant led to the isolation of 3-(3-methylbut-2-en-1-yl)isoguanine¹) (1), a new purine derivative, and 19-hydroxyspruceanol 19- $O-\beta$ -D-glucopyranoside¹) (2), a new cleistanthane-type diterpene glucoside, as well as of eight known compounds, including one lignan glycoside, mananthoside I [8], one polyphenol, (-)-epigallocatechin [9], four aromatic compound glucosides, isotachioside [10], carthamoside B₅ [11], hovetrichoside A [12], and 3,4-dihydroxyphenylpropanol 3-O- β -D-glucopyranoside [13], and two megastigmane glycosides, turpenionosides A and B [14]. The above compounds are reported from this plant for the first time. In this paper, we describe the isolation and structural elucidation of 1 and 2.

Results and Discussion. – Compound **1** showed *quasi*-molecular-ion peaks at m/z 220 ($[M + H]^+$), 242 ($[M + Na]^+$), and 218 ($[M - H]^-$) in the positive-ion- and negative-ion-mode ESI-MS, resp., in accord with the molecular formula $C_{10}H_{13}N_5O$, which was confirmed by the HR-ESI-MS. The structure of **1** was elucidated to be 3-(3-methylbut-2-en-1-yl)isoguanine¹) on the basis of NMR analyses. The ¹H-NMR spectrum (*Table 1*) showed a *s* of an aromatic H-atom at $\delta(H)$ 8.11, and signals of a

¹⁾ Trivial atom numbering; for systematic names, see Exper. Part.

^{© 2010} Verlag Helvetica Chimica Acta AG, Zürich



3-methylbut-2-en-1-yl group at δ (H) 5.26 (t, J = 6.8 Hz, 1 H), 4.54 (d, J = 6.8 Hz, 2 H), 1.78 (s, 3 H), and 1.66 (s, 3 H). The ¹³C-NMR and DEPT spectra (*Table 1*) displayed ten C-atom signals. Among them, five C-atoms were attributed to the (3-methylbut-2-en-1-yl) unit (δ (C) 135.7, 119.0, 40.5, 25.3, and 17.9); the remaining four sp² quaternary C-atoms (δ (C) 151.5, 150.6, 103.1, and 150.9), and one sp² CH group (δ (C) 142.0) combined with five N-atoms formed a skeleton of isoguanine [15], indicating an alkenylated isoguanine. The HMBC experiments (*Table 1*) confirmed the isoguanine unit and the 3-methylbut-2-en-1-yl group attached at N(3) by the cross-peaks H–C(8)/C(4) and C(5), as well as CH₂(10)/C(2) and C(4).

Table 1. ¹H- and ¹³C-NMR Data (400 and 100 MHz, resp.; (D_6)DMSO) of 1^1). δ in ppm, J in Hz.

	$\delta(C)$	$\delta(H)$	HMBC $(H \rightarrow C)$		$\delta(C)$	$\delta(H)$	HMBC $(H \rightarrow C)$
C(2)	151.5 (s)			CH ₂ (10)	40.5 (t)	4.54 (d, J = 6.8)	C(2), C(4), C(12)
C(4)	150.6 (s)			H - C(11)	119.0 (d)	5.26 (t, J = 6.8)	C(13), C(14)
C(5)	103.1(s)			C(12)	135.7 (s)		
C(6)	150.9 (s)			Me(13)	17.9(q)	1.78(s)	C(11), C(14)
H-C(8)	142.0 (<i>d</i>)	8.11 (s)	C(4), C(5)	Me(14)	25.3 (q)	1.66 (s)	C(11), C(13)

Compound 2 was obtained as a white amorphous powder. Its molecular formula was determined to be $C_{26}H_{38}O_8$ by the HR-ESI-MS. Acid hydrolysis of 2 gave a Dglucose as sugar moiety. The structure of 2 was established to be 19-hydroxyspruceanol 19-O- β -D-glucopyranoside¹) by interpretation of its spectroscopic parameters and comparison with those of spruceanol (=(2R,4aR,10S)-8-ethenyl-1,2,3,4,4a,9,10,10aoctahydro-1,1,4a,7-tetramethylphenanthrene-2,6-diol) [16]. The ¹H- and ¹³C-NMR and HMQC spectra of 2 (Table 2) showed that the aglycone contained 20 C-atoms and 25 C-bearing H-atoms (3 Me, 6 CH₂, 4 CH, and 7 C). Among which a pentasubstituted aromatic ring (δ (C) 154.9, 148.8, 140.4, 125.1, 121.1, and 111.2 (d); δ (H) 6.64 (s)) and attached Me group ($\delta(C)$ 13.8 (q); $\delta(H)$ 2.09 (s)), an ethenyl group ($\delta(C)$ 137.8 (d) and 119.9 (t); $\delta(H)$ 6.57, 5.46, and 5.05 (each dd, ${}^{2}J = 2.0$, ${}^{3}J = 17.9$, 11.4 Hz)), one isolated O-bearing CH₂ group (δ (C) 72.8; δ (H) 4.30 and 3.57 (each d, J = 10.2 Hz)), one Obearing CH group (δ (C) 80.8; δ (H) 3.29 (dd, J = 11.3, 5.9 Hz)), and two tertiary Me groups (δ (H) 1.24 and 1.22) were assigned, indicating a spruce analogue [16]. Comparison of the ¹³C-NMR data of the aglycone of **2** with those of spruceanol revealed that the most important difference was the isolated O-bearing CH_2 group of 2 instead of a Me group ($\delta(C)$ 15.4) for Me(19) of spruce anol, indicating that the

aglycone of **2** was 19-hydroxyspruceanol. The HMBC spectrum (*Table 2*) exhibited cross-peaks between $CH_2(19)$ and C(3), C(4), C(5), C(18) and C(1'), establishing that **2** is 19-hydroxyspruceanol 19-O- β -D-glucopyranoside.

	$\delta(C)$	$\delta(\mathrm{H})$	HMBC $(H \rightarrow C)$
CH ₂ (1)	39.5 (t)	2.27 (dt , $J = 13.8, 3.3, H_a$),	C(3), C(5), C(10)
		1.45 (td, $J = 13.6, 3.2, H_{\beta}$)	C(5), C(10), C(20)
$CH_{2}(2)$	29.7 (t)	1.79 (br. $d, J = 13.4, H_a$),	
		1.96 $(qd, J = 13.0, 3.2, H_{\beta})$	
H-C(3)	80.8(d)	3.29 (dd, J = 11.3, 5.9)	C(4), C(18), C(19)
C(4)	44.2 (s)		
H-C(5)	52.7 (<i>d</i>)	1.32 (dd, J = 12.3, 2.0)	C(3), C(4), C(6), C(7), C(9), C(10), C(18), C(19), C(20)
CH ₂ (6)	21.6(t)	$1.73 - 1.79 (m, H_a),$	C(5), C(7), C(10)
2.		1.98 (br. $d, J = 12.7, H_{\beta}$)	C(4), C(5), C(7), C(8), C(10)
$CH_{2}(7)$	31.6 (<i>t</i>)	$2.74 (dd, J = 16.9, 5.2, H_a),$	C(6), C(8), C(9), C(14)
		2.45 (ddd , $J = 16.9, 11.5, 7.0, H_{\beta}$)	C(5), C(6), C(8), C(9), C(14)
C(8)	125.1(s)		
C(9)	148.8 (s)		
C(10)	39.2 (s)		
H–C(11)	111.2 <i>(d)</i>	6.64 (<i>s</i>)	C(8), C(9), C(10), C(12), C(13), C(17)
C(12)	154.9(s)		
C(13)	121.1(s)		
C(14)	140.4(s)		
C(15)	137.8 (d)	6.57 (dd, J = 17.9, 11.4)	C(8), C(13), C(14)
$CH_{2}(16)$	119.9 (t)	5.46 (dd, J = 11.4, 2.0),	C(14)
		5.05 (dd, J = 17.9, 2.0)	C(14), C(15)
Me(17)	13.8 (q)	2.09(s)	C(8), C(9), C(11), C(12), C(13), C(14)
Me(18)	24.4(q)	1.24 (s)	C(1), C(5), C(9), C(10), C(19)
$CH_{2}(19)$	72.8(t)	3.57 (d, J = 10.2), 4.30 (d, J = 10.2)	C(3), C(4), C(5), C(18), C(1')
Me(20)	26.3(q)	1.22 (s)	C(1), C(5), C(9), C(10)
H-C(1')	105.5(d)	4.22 (d, J = 7.9)	C(19), C(2'), C(5')
H-C(2')	75.5(d)	3.19(t, J = 8.3)	
H-C(3')	78.6(d)	3.36(t, J = 8.5)	
H-C(4')	72.0(d)	3.30(t, J = 8.5)	
H-C(5')	78.4(d)	3.26 (br. $dd, J = 8.5, 4.7$)	
CH ₂ (6')	63.1 (<i>t</i>)	3.86 (dd, J = 12.0, 1.4),	
		3.68 (dd, J = 12.0, 4.7)	

Table 2. ¹H- and ¹³C-NMR Data (400 and 100 MHz, resp.; CD₃OD) of **2**¹). δ(H) in ppm, J in Hz.

This study was supported by grants from the Key New Drug Creation and Manufacturing Program (2009ZX09301-001) of the National Science & Technology Major Project of the Ministry of Science & Technology of China.

Experimental Part

General. Column chromatography (CC): silica gel (SiO₂; 200–300 mesh; Qingdao Haiyang, Co., Ltd., P. R. China), Sephadex LH-20 (Pharmacia Biotech AB, Uppsala, Sweden), ODS-A gel (Mitsubishi Chemical Industries Co., Ltd., Japan). TLC: silica gel HSGF₂₅₄ (Yantai Jiangyou Guijiao Kaifa Co., Ltd.,

P. R. China). Optical rotation: *Perkin-Elmer-341* polarimeter. UV Spectra: *Shimadzu-UV-2550* spectrophotometer; λ_{max} (log ε) in nm. IR Spectra: *Nicolet-Magna-750-FTIR* spectrometer; KBr pellets; in cm⁻¹. NMR Spectra: *Bruker-AV-400* instrument at 400 (¹H) and 100 MHz (¹³C); in (D₆)DMSO or CD₃OD; δ in ppm rel. to Me₄Si; *J* in Hz. ESI-MS and HR-ESI-MS: *Bruker-Esquire-3000-plus* and *Finnigan-LC-QDECA* mass spectrometers, resp.; in *m/z* (rel. int.).

Plant Material. The fresh whole plants of *P. reticulatus* were collected in the Yongning County, Guangxi Province, P. R. China. The plant was identified by Prof. *S.-J. Wei* of the Guangxi Traditional Chinese Medicine University. A voucher specimen (No. 09-108) is deposited at the Herbarium of the Shanghai Institute of Materia Medica.

Extraction and Isolation. Air-dried and powdered whole plants of *P. reticulates* (5 kg) were extracted three times with 201 of 75% EtOH at r.t. The concentrated extract was partitioned between H₂O and petroleum ether, CHCl₃, AcOEt, and BuOH, resp. The BuOH fraction (108 g) was subjected to CC (SiO₂ (2 kg), column i.d. 10×90 cm, CHCl₃, CHCl₃/MeOH 100:1, 50:1, 20:1, 10:1, 6:1, and 3:1): *Frs. A* – *G. Fr. E* yielded a solid which was further purified by CC (*Sephadex LH-20*, MeOH): **1** (12 mg). *Fr.* f (10.9 g) was separated by CC (SiO₂, CHCl₃/MeOH 10:1, 8:1, 6:1, and 3:1): *Frs. A* – *G. Fr. E* yielded a solid which was further purified by CC (*Sephadex LH-20*, MeOH): **1** (12 mg). *Fr.* f (10.9 g) was separated by CC (SiO₂, CHCl₃/MeOH 10:1, 8:1, 6:1, and 3:1): *Frs. F*₁–*F*₄. From *Fr. F*₂ (320 mg), **2** (15 mg) was obtained after two CC (1. *ODS-A* gel, 20%, 50%, 70% and 95% MeOH/H₂O; 2. Foregoing 70% portion, *Sephadex LH-20*, MeOH). *Fr. F*₃ (550 mg) was subjected to CC (SiO₂, CHCl₃/MeOH/H₂O 5:1:0.1): *Frs. F*_{3.1} – *F*_{3.8}. *Fr. F*_{3.1} (45 mg) gave isotachioside (4 mg) after purification by two CC (1. *ODS-A* gel, 10–50% MeOH/H₂O; 2. *Sephadex LH-20*, MeOH). By the same procedure as applied to *Fr. F*_{3.1}, *Frs. F*_{3.4} (62 mg), *F*_{3.5} (80 mg), *F*_{3.6} (50 mg), *F*_{3.7} (45 mg), and *F*_{3.8} (60 mg) afforded (–)-epigallocatechin (13 mg), mananthoside I (32 mg), carthamoside B₅ (14 mg), hovetrichoside A (4 mg), and 3,4-dihydroxyphenylpropanol 3-*O-β*-D-glucopyranoside (6 mg), as well as turpenionosides A (8 mg) and B (12 mg), resp.

Acid Hydrolysis of 2. A soln. of 2 (2 mg) in 2M HCl/dioxane 1:1 (2 ml) was refluxed for 2 h. After cooling, the soln. was neutralized with NaHCO₃ and then filtrated to remove the solid. The filtrate was subjected to CC (*Sephadex LH-20*, 50% MeOH/H₂O) to afford a sugar fraction. This sugar fraction and standard D-glucose (*Sigma*, USA) were each treated with L-cysteine methyl ester hydrochloride (2 mg) in pyridine (1 ml) at 60° for 1 h. Then, the soln. was treated with *N*,*O*-bis(trimethylsilyl)trifluoroacetamide (0.02 ml) at 60° for 1 h. Subsequently, the supernatant was subjected to GC analysis (*Supelco*, 230°, flow rate 15 ml/min): D-Glucose (t_R 24.2 min) was detected.

3-(3-Methylbut-2-en-1-yl)isoguanine (=6-Amino-3,9-dihydro-3-(3-methylbut-2-en-1-yl)-2H-purin-2-one; **1**): White amorphous powder. UV (MeOH): 203 (5.36), 291 (5.08). IR: 3325, 1767, 1660, 1604, 1576, 1493, 1448, 1410, 1302, 756, 552. ¹H- and ¹³C-NMR and HMBC: *Table 1*. ESI-MS (pos.): 220 ($[M + H]^+$), 242 ($[M + Na]^+$), 439 ($[2 M + H]^+$). ESI-MS (neg.): 218 ($[M - H]^-$), 437 ($[2 M - H]^-$). HR-ESI-MS: 242.1021 ($[M + Na]^+$, C₁₀H₁₃N₅NaO⁺; calc. 242.1018).

19-Hydroxyspruceanol 19-O-β-D-Glucopyranoside (=rel-[(1R,2R,4aR,10aS)-8-Ethenyl-1,2,3,4, 4a,9,10,10a-octahydro-2,6-dihydroxy-1,4a,7-trimethylphenanthren-1-yl]methyl β-D-Glucopyranoside; **2**): White amorphous powder. $[a]_{D}^{25} = -42$ (c = 0.18, MeOH). UV (MeOH): 211 (4.95). IR: 3421, 1632, 1425, 1398, 1269, 1078, 1040. ¹H- and ¹³C-NMR and HMBC: *Table 2*. ESI-MS (pos.): 501 ($[M + Na]^+$). ESI-MS (neg.): 523 ($[M + HCOO^{-}]^{-}$), 955 ($[2 M - H]^{-}$). HR-ESI-MS: 501.2469 ($[M + Na]^+$, $C_{26}H_{38}NaO_{3}^+$; calc. 501.2464).

REFERENCES

- The Health Bureau of Guangxi Province (Ed.), 'Compilation of Medicinal Herbs in Guangxi, II', Guangxi People Press, Nanning, 1974, p. 1588.
- [2] G. H. Schmelzer, A. Gurib-Fakim, R. Arroo, C. H. Bosch, A. de Ruijter, M. S. J. Simmonds, 'Plant Resources of Tropical Africa 11(1) – Medicinal Plants 1', Backhuys Publishers, Wageningen, Netherlands, 2008.
- [3] E. Omulokoli, B. Khan, S. C. Chhabra, J. Ethnopharmacol. 1997, 56, 133.
- [4] S. Kumar, D. Kumar, R. R. Deshmukh, P. D. Lokhande, S. N. More, V. D. Rangari, *Fitoterapia* 2008, 79, 21.

- [5] T. Begum, M. S. Rahman, M. A. Rashid, Dhaka Univ. J. Pharm. Sci. 2006, 5, 21.
- [6] B. K. Das, S. Bepary, B. K. Datta, A. A. Chowdhury, M. S. Ali, A. S. Rouf, *Pak. J. Pharm. Sci.* 2008, *21*, 333.
- [7] S.-H. Lam, C.-Y. Wang, C.-K. Chen, S.-S. Lee, Phytochem. Anal. 2007, 18, 251.
- [8] J.-M. Tian, H.-P. He, Y.-T. Di, X.-W. Yang, Z.-L. Gao, X.-J. Hao, J. Asian Nat. Prod. Res. 2008, 10, 228.
- [9] S. Valcic, J. A. Burr, B. N. Timmermann, D. C. Liebler, Chem. Res. Toxicol. 2000, 13, 801.
- [10] X.-N. Zhong, H. Otsuka, T. Ide, E. Hirata, Y. Takeda, Phytochemistry 1999, 52, 923.
- [11] Y.-Z. Zhou, H. Chen, L.Qiao, X. Lu, H.-M. Hua, Y.-H. Pei, Helv. Chim. Acta 2008, 91, 1277.
- [12] K. Yoshikawa, N. Mimura, S. Arihara, J. Nat. Prod. 1998, 61, 1137.
- [13] T. Ishikawa, Y. Sega, J. Kitajima, Chem. Pharm. Bull. 2001, 49, 840.
- [14] Q. Yu, H. Otsuka, E. Hirata, T. Shinzato, Y. Takeda, Chem. Pharm. Bull. 2002, 50, 640.
- [15] J.-W. Chern, H.-Y. Lee, M. Huang, F.-J. Shish, Tetrahedron Lett. 1987, 28, 2151.
- [16] R. W. Denton, W. W. Harding, C. I. Anderson, H. Jacobs, S. McLean, W. F. Reynolds, J. Nat. Prod. 2001, 64, 829.

Received April 28, 2010